Effect of Reduced Aortic Compliance on Cardiac Efficiency and Contractile Function of In Situ Canine Left Ventricle

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This study tests the hypothesis that arterial vascular stiffening adversely influences in situ left ventricular contractile function and energetic efficiency. Ten reflex-blocked anesthetized dogs underwent a bypass operation in which a Dacron graft was sewn to the ascending aorta and connected to the infrarenal abdominal aorta via a plastic conduit. Flow was directed through either native aorta or plastic conduit by placement of vascular clamps. Arterial properties were measured from aortic pressure–flow data, and ventricular function was assessed by pressure–volume (PV) relations. Coronary sinus blood was drained via an extracorporeal circuit for direct measurement of myocardial O2 consumption (MVO2). Data at multiple steady-state preload volumes were combined to derive chamber function and energetics relations. Energetic efficiency was assessed by the inverse slope of the MVO2–PV area relation. Directing flow through plastic versus native aorta resulted in a 60–80% reduction in compliance but little change in mean resistance. Arterial pulse pressure rose from 34 to 99 mm Hg (p < 0.001). Contractile function assessed by the end–systolic PV relation, stroke work–end-diastolic volume relation, and dP/dtmax at matched end-diastolic volume did not significantly change. However, MVO2 increased by 32% (p < 0.01) and was matched by a rise in PV area, such that the MVO2–PV area relation and efficiency was unaltered. The MVO2 required to sustain a given stroke volume, however, increased from 20% to 40%, depending on the baseline level (p < 0.001). Thus, whereas the contractile function and efficiency of normal hearts are not altered by ejection into a stiff vascular system, the energetic cost to the heart for maintaining adequate flow is increased. This suggests a mechanism whereby human vascular stiffening may yield little functional decrement at rest but limit reserve capacity under conditions of increased demand. (Circulation Research 1992;71:490–502)

KEY WORDS • aortic input impedance • ventriculovascular coupling • myocardial energetics • pressure–volume relations • ventricular function

Human aortas undergo progressive degeneration due to normal aging1–7 and disease processes such as chronic hypertension.8,9 The dominant change is that of reduced vascular distensibility, which has an impact on measures of arterial properties such as the shape of the arterial pressure and flow wave,5,10 aortic input impedance spectra,5,8,11 and pulse-wave velocity.7,9,12 Although it is widely believed that these abnormalities of vascular load have adverse effects on cardiac contractile function and energetic efficiency,12–14 this hypothesis has remarkably little direct supporting data.

For example, isolated canine heart studies have reported little change in the end-systolic pressure–volume relation (ESPVR) despite twofold changes in total compliance, characteristic impedance, and resistance.15 Myocardial efficiency, defined by the relation between total pressure–volume work (pressure–volume area [PVA]) and oxygen consumption, has also been shown to be little influenced by vascular load or ejection history.16–18 We recently reported that lowering vascular resistance could have positive effects on both contractility and efficiency19; however, the relevance of this finding to in situ hearts ejecting into a stiff vasculature is unknown. Other prior studies have attempted to measure in situ effects of aortic stiffening on cardiac function and energetics by replacing the aorta with an artificial conduit or by physically stiffening the native aorta.20–23 However, these data largely focused on vascular properties, providing little direct assessment of ventricular performance or energetics. Furthermore, none of these studies demonstrated increased myocardial oxygen consumption (MVO2) from ejection into a stiffened aorta.22,23 Thus, despite the popularity of the notion that vascular stiffening adversely affects cardiac contractility and energetics, confirmation with precise measurements of the magnitude of this interaction has not been made.

The purpose of the present study was to determine in situ whether acutely reduced vascular compliance sig-
nificantly alters ventricular contractility, M\textsubscript{VO}\textsubscript{2}, or efficiency. Pressure–volume analysis was used to assess chamber properties and energetics. A novel surgical preparation was developed in which virtually the entire canine thoracic aorta was bypassed by a stiff plastic conduit and the ventricular outflow was directed through either the native or stiff aortas. Total M\textsubscript{VO}\textsubscript{2} was directly measured and compared with simultaneous pressure–volume data for efficiency calculations. These data provide the first direct in situ evidence confirming energetic consequences but insignificant systolic contractile effects from cardiac ejection into a stiff vascular system.

**Materials and Methods**

**Preparation**

Ten adult mongrel dogs (30–35 kg) were anesthetized with pentobarbital (30 mg/kg i.v.). The dogs were pretreated with 100 mg i.m. hydrocortisone and 50 mg p.r. indomethacin before study, which served to stabilize the preparation during extracorporeal blood circulation. The animals were intubated and ventilated on a volume respirator with 40% enhanced inspired oxygen. Ventilation was guided by arterial blood gases obtained every 30–60 minutes.

Both carotid and femoral arteries were cannulated by fluid-filled catheters to allow arterial blood sampling and pressure monitoring and to provide a route for cerebral perfusion after aortic bypass. The right femoral vein was cannulated for intravenous fluid, blood, and drug administration. The animals underwent a midsternotomy and midline abdominal incision. The inferior and superior venae cavae and the descending thoracic aorta at the level of the diaphragm were snared with umbilical ties. The pericardium was opened, and the heart was suspended in a pericardial cradle (Figure 1). The proximal aortic root was carefully dissected free from surrounding connective and fatty tissue, and the right brachiocephalic artery was tied off and divided. A 6F micromanometer catheter (presoaked for 2 hours at room temperature and calibrated to a mercury manometer) was inserted through a purse-string suture in the high left ventricular free wall to monitor intracavity pressure (site A in Figure 1). Distal (femoral artery) perfusion pressure was monitored using a fluid-filled line connected to a Statham gauge.

The proximal ascending aorta was partially occluded with an aortic clamp at or just proximal to the origin of the brachiocephalic artery. This isolated a portion of the aortic wall (site B) onto which a 2–3-cm long Dacron vascular graft was sewn by end-to-side anastomosis using a pair of running 4–0 prolene sutures. With the aorta appropriately clamped, there was minimal rise in left ventricular pressure, and distal perfusion was well maintained. The Dacron graft (18 or 14 mm o.d.) was matched to the size of the proximal aorta. After partial unclamping of the aortic wall to ensure adequate hemostasis, the suture line was stabilized with felt pledgets and collagen fiber (Avitene, MedChem Prod. Inc., Woodburn, Mass.), and the aorta was left partially clamped to enable a thrombotic seal to form at the anastomosis.

The abdominal aorta was then dissected free from surrounding tissue, cross-clamped just above the iliac bifurcation, and partially transected to allow insertion of a short T-tube cannula (site C in Figure 1). This was secured in place with umbilical tape. Heparin (5,000 units) was administered intravenously. The common port of the T-tube cannula was clamped off, distal perfusion was restored, and the abdomen was closed with surgical clamps.

A purse-string suture was placed at the left ventricular apex, and an 8F multielectrode conductance catheter was advanced through the apex and extended to a position just above the aortic valve (by monitoring intraluminal pressure) (site D in Figure 1). The catheter was attached to a signal generator/processor (Sigma V, Lecoom, The Netherlands), which provided an excitation current at base and apical electrodes and measured voltages between intervening electrode pairs along the catheter. These voltages were related to chamber blood volume as previously described.\textsuperscript{24,25} Examination of individual pressure–volume segment loops determined which electrodes were intracavity (counterclockwise loop), and electronic switching (i.e., eliminating segments within the aortic root from the total volume calculation) enabled the appropriate electrodes to be used. Once the catheter was correctly positioned, it was secured by the apical purse-string suture.
An ultrasonic flow probe (Transonics, Ithaca, N.Y.) (site E in Figure 1) was placed around the aortic root within 2 cm of the aortic valve and immediately below the Dacron anastomosis (site B). A 6F micromanometer catheter was inserted via the left brachiocephalic artery (site F) and secured in place so that the catheter tip lay just downstream from the aortic flow probe. Since both aortic arch branches were now ligated, head perfusion was maintained by flow diverted from the abdominal aorta using a perfusion line connecting a side port of the abdominal aortic cannula to the carotid cannula.

The right atrial appendage was cannulated with a modified 14F Foley (urinary drainage) catheter by using a thin slightly angled metal obturator to assist passage. The catheter was advanced into the proximal coronary sinus (CS), and the balloon was inflated with 3–4 ml saline so that it fit snugly in the CS orifice (site G). The catheter was secured in place with a proximal tie around the right atrial appendage. With the metal obturator removed, CS blood was allowed to drain by gravity through an in-line ultrasound volume flow probe (site H) (Transonics), and a portion of the CS blood was diverted to a blood oxygen content difference analyzer (site I). Residual CS blood (plus outflow from the blood analyzer) was drained by gravity into a reservoir (site J). The reservoir blood was pumped by peristaltic pump through a heat exchanger (37°C), air trap, and arterial filter (site K) and returned to the right femoral vein. A custom electronic switching device detected the upper fluid level of blood in the reservoir and provided feedback control to the pump. The fluid level could be raised or lowered to allow changes in steady-state venous return and cardiac preload. After completion of the CS drainage circuit, an additional 5,000 units heparin was administered.

The distal abdominal and proximal aortic cannulas were connected by a 58-cm (0.5-in.-i.d., 0.625-in.-o.d.) plastic tube (Tygon, Norton, Akron, Ohio) with a compliance of 3×10⁻⁷ ml/mm Hg over the physiological pressure range. Any residual air was removed from the tube, and then the proximal aortic clamp was removed to allow flow to proceed in parallel through both the native aorta and Tygon conduit. After establishing the stability of the preparation, the plastic conduit was then reclamped both proximally and distally to provide native aortic flow only.

Protocol

At least 10 minutes was allowed for stabilization after the preparation was completed. Pharmacological support of blood pressure and contractility was required in some dogs and was provided by low dose infusion of epinephrine (1–3 µg/kg per minute). Reflex blockade was achieved by hexamethonium HCl (10 mg/kg) and assessed by lack of heart rate change with varying load and transient cessation of carotid flow. Additional hexamethonium (5 mg/kg) was administered if required.

Left ventricular outflow was directed through either the native aorta or Tygon conduit by placement of vascular occlusion clamps. Flow through the native aorta was achieved by clamping off the Tygon conduit at the proximal and distal connections. Flow through the Tygon conduit was obtained by clamping the native aorta immediately beyond the proximal anastomosis (mid ascending aorta) and at the diaphragm. This latter site was chosen to avoid hemodynamic effects of renal or bowel ischemia. Multiple steady-state pressure–volume, aortic pressure–flow, coronary sinus flow, and arteriovenous oxygen difference data were obtained at varying preload volumes. Preload was reduced in gradual steps by slowing the speed of the pump returning blood from the CS drainage reservoir (site K in Figure 1). With each load change, all variables were allowed to equilibrate for 1–2 minutes before recording. Once a set of data (four to seven preload volumes) was obtained for either the native aorta or Tygon conduit, the clamps were repositioned so that flow was directed via the alternate route. After several minutes for stabilization, the protocol and measurements were repeated. In addition to steady-state data, pressure–volume loop data were also obtained during acute transient preload reduction by applying traction to inferior and superior vena caval snare.

Data were monitored on an eight-channel chart recorder and displayed with custom-developed data acquisition/display software using an Intel 286 processor–based microcomputer. Data were digitally recorded at 200 Hz and stored on removable hard drives for subsequent analysis.

At the end of the study, the animals were euthanized by pentobarbital overdose and potassium chloride cardioplegia. The CS drainage catheter placement and balloon seal were assessed by injection of monastral blue dye retrogradely down the CS catheter lumen (before cardioplegia) to test for leakage into the right atrium and by direct postmortem inspection. The heart was removed from the chest, and right and left ventricles (free wall plus septum) were weighed.

Volume Signal Calibration

Conductance catheter volumes were calibrated to the aortic flow probe by determining the ratio of integrated flow to catheter-derived stroke volume (SV). This gain was calculated for each steady-state beat (coefficient of variation, 6.2%) in a given preload run, and the results were averaged to obtain a mean calibration gain. Variation in mean gain between native and Tygon data runs averaged 11%; however, this as well as interanimal differences did not affect the results, since a new mean gain was determined and applied to each preload data set. The conductance signal also includes an offset, which is primarily due to cardiac muscle conductance. Estimation of this offset was not made in the present study, since the focus was on relative changes due to rapidly altered vascular properties in the same heart. Furthermore, the primary function parameters such as stroke work (SW), SV, and PVA are not influenced by a volume offset.

Data Analysis

Ventricular systolic function. Data from multiple steady-state preloads were combined to determine systolic and diastolic pressure–volume relations. Left ventricular contractile function was assessed by ESPVR, the SW–end-diastolic volume (Vd) relation, and the maximal rate of pressure rise (dP/dtm.,) at a common Vd. The ESPVR was fit by linear regression (using an iterative algorithm) to obtain a slope and volume intercept (V0). SW was the digitally integrated area of each pressure–volume loop and was plotted versus Vd.
(volume at the lower right corner of the loop) to generate the SW–V_{ed} relation. These relations were highly linear and fit by least-squares regression. The time derivative of ventricular pressure was digitally calculated by running a weighted five-point slope. \(dp/dt_{max}\) was compared for native versus Tygon aortas at a matched \(V_{ed}\) to minimize loading effects.

**Myocardial energetics.** Myocardial energetics was assessed by analysis of the relation between left ventricular MVO_{2} per beat and total pressure–volume work (PVA) as described by Suga. We also examined the relation between MVO_{2} per beat and cardiac SV to assess the oxygen cost of maintaining or increasing SV (or cardiac output) while ejecting into a stiffer arterial system. Over a physiological range of mean vascular pressures, cardiovascular reserve capacity is closely tied to the ability of the system to enhance cardiac output, making this relation important.

MVO_{2} was calculated from the product of coronary sinus blood flow and arteriovenous oxygen difference. PVA is the sum of SW plus the area bounded by the ESPVR and diastolic pressure–volume relations between end-systolic volume and \(V_e\). \(V_e\) was determined by linear extrapolation of the ESPVR, with an average standard error of the estimate of \(\pm 3.1\) mL. To obtain the non-SW portion of PVA, the diastolic data were fit by nonlinear regression, and the area between diastolic and systolic pressure–volume relations was determined by analytical integration. MVO_{2}–PVA relations were fit by linear regression. The slope provides a measure of chemomechanical transduction efficiency, and the offset provides O_{2} consumption for basal metabolism and excitation–contraction coupling.

This study examined effects of altered arterial load on left ventricular performance; therefore, total MVO_{2} was adjusted to reflect left ventricular MVO_{2} only. This was done by assuming that the MVO_{2} values in right and left ventricles were proportional to their masses (linking the septum with the left ventricle). Thus, MVO_{2} was multiplied by the ratio of left ventricular/(left ventricular+right ventricular) weight. Finally, to compare MVO_{2}–PVA relations between hearts, both MVO_{2} and PVA were normalized to 100 g left ventricular weight as described by Suga.

**Vascular properties.** Arterial properties were assessed from ascending aortic pressure–flow measurements. Impedance spectra from steady-state beats were obtained by discrete Fourier series analysis. Total mean resistance was the zero-order term from the impedance spectra, and characteristic impedance was the average modulus from 5 to 12 Hz. Flow signal noise was assessed by assuming a zero flow level during late diastole and determining the amplitude of fluctuations during this period (by Fourier analysis). Frequency coefficients that did not exceed this level were not included in the analysis. Peripheral resistance based on a three-element Windkessel model was estimated as the difference between total mean resistance and characteristic impedance.

Total vascular compliance was estimated using the method of Liu et al. This approach, also based on the Windkessel model, calculates arterial compliance from the area under the diastolic pressure waveform rather than from nonlinear fits. Thus, total vascular compliance equals \(A_{21}/R_{a}(P_{1}–P_{2})\) (Equation 11 in Reference 27), where \(A_{21}\) is the area under the diastolic pressure wave between two arbitrary time points (1 and 2), and \(P_{1}–P_{2}\) is the corresponding pressure difference. Since diastolic waveforms rarely followed a pure monoeponential decay and also displayed wave oscillations, particularly in native aorta data, total vascular compliance estimates varied with the portion of diastole (position of time points 1 and 2) used in this calculation. An upper range estimate was obtained using the entire diastolic period (i.e., \(P_{1}\) is incisura pressure, and \(P_{2}\) is end-diastolic pressure). A lower range estimate was obtained by positioning \(P_{1}\) at the peak of any reflected wave, so that only a monotonic declining portion remained. Both estimates are reported.

In addition to steady-state data, a subset of hearts underwent random pacing to obtain broadband impedance spectra. Atrial pacing leads were attached, and a random number generator was used to obtain cycle lengths between 250 and 1,200 msec. PACing data were continuously recorded at 100 Hz for 4 minutes. Data were divided into eight contiguous bins (2,048 points, or 10 seconds per bin), windowed using a Hanning filter, analyzed by fast Fourier transform, and averaged in the frequency domain to yield impedance spectra. Modulus, phase, and coherence were determined at each frequency.

**Statistical analysis.** All data are presented as mean±SD. Steady-state variables were compared by paired t tests, with statistical significance accepted at \(p<0.05\). Linear regression slopes and offsets for the various relations (e.g., end-systolic pressure versus end-systolic volume; MVO_{2} versus PVA; SW versus \(V_{ed}\)) were also compared by paired t tests. In addition, all of the raw data for each relation was combined into a single multiple-regression model to provide a more robust assessment for differences. The model included dummy variables coding for the aorta type and for variability of slope and offset among individual dogs. The output provided the mean relation for both the native and Tygon aorta and a determination of whether aorta type significantly altered either mean slope or offset. Numerical analysis was performed on an Intel 386 processor-based microcomputer using custom-designed software and a commercial statistics package (SYSTAT, Inc., Evanston, Ill.).

**Results**

**Aortic Impedance and Pressure–Flow and Pressure–Volume Contours**

Diversion of flow from the native aorta to the Tygon conduit resulted in a dramatic increase in several measures of pulsatile arterial load (Table 1). Arterial systolic pressure and pulse pressure increased by 51.8 mm Hg (47%) and 64.8 mm Hg (190%), respectively (both \(p<0.01\)). Arterial diastolic pressure decreased slightly, yielding a net +11 mm Hg increase in mean pressure. Cardiac output was not significantly altered. Data provided in Table 1 were obtained at steady state, with the blood return flow rate adjusted to maintain a similar volume level in the external reservoir.

Figure 2 displays an example of aortic pressure and flow waveforms and ventricular pressure–volume loops for flow through native and Tygon “aortas,” as well as intermediate combinations. The examples on the left show flow through the native aorta only, revealing a narrow pulse pressure, rounded arterial waveform, early
TABLE 1. Mechanical, Vascular, and Energetic Consequences of Switch to Stiff (Tygon) Aorta

<table>
<thead>
<tr>
<th>Variable</th>
<th>Native aorta</th>
<th>Tygon conduit</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vascular</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Psys (mm Hg)</td>
<td>109.8±18.6</td>
<td>161.6±34.9</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Pdia (mm Hg)</td>
<td>75.7±18.1</td>
<td>62.7±17.9</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Pmean (mm Hg)</td>
<td>91.7±20.5</td>
<td>102.6±22.5</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>PP (mm Hg)</td>
<td>34.1±8.3</td>
<td>98.9±24.8</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Rf (mm Hg·mL⁻¹·sec⁻¹)</td>
<td>3.04±1.5</td>
<td>3.66±1.05</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>Rc (mm Hg·mL⁻¹·sec⁻¹)</td>
<td>0.27±0.11</td>
<td>0.41±0.17</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>Rs (mm Hg·mL⁻¹·sec⁻¹)</td>
<td>2.77±1.4</td>
<td>3.24±0.92</td>
<td>NS</td>
</tr>
<tr>
<td>Ca (ml/mm Hg)</td>
<td>1.65±0.85–0.63±0.38*</td>
<td>0.19±0.07–0.11±0.08*</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Ventricular</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pes (mm Hg)</td>
<td>104.7±23.1</td>
<td>153.4±31.5</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Pea (mm Hg)</td>
<td>6.8±2.4</td>
<td>9.4±3.8</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>CO (l/min)</td>
<td>1.9±0.55</td>
<td>1.7±0.44</td>
<td>NS</td>
</tr>
<tr>
<td>EF (%)</td>
<td>50.4±12.5</td>
<td>42.1±13.4</td>
<td>NS</td>
</tr>
<tr>
<td>HR (bpm)</td>
<td>119.5±9.8</td>
<td>121.1±12.8</td>
<td>NS</td>
</tr>
<tr>
<td>SW (mm Hg·ml)</td>
<td>1,572.8±482.4</td>
<td>1,630.4±425.3</td>
<td>NS</td>
</tr>
<tr>
<td>Vaved–Ves (ml)</td>
<td>35.6±14.5</td>
<td>40.2±18.3</td>
<td>0.18</td>
</tr>
<tr>
<td>Va–Ves (ml)</td>
<td>19.1±11.3</td>
<td>25.9±15.1</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>Energetics</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>MV02 (ml O2·100 g⁻¹·beat⁻¹)</td>
<td>0.102±0.03</td>
<td>0.135±0.049</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>PVA (mm Hg·ml/100 g)</td>
<td>2,498.8±863.7</td>
<td>3,450.1±1,547.3</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>EFF</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>SW/MV02 (%)</td>
<td>22.9±10.9</td>
<td>17.6±5.4</td>
<td>0.052</td>
</tr>
<tr>
<td>1/α (%)</td>
<td>35.5±11.2</td>
<td>34.4±9.1</td>
<td>NS</td>
</tr>
</tbody>
</table>

Psys, Pdia, and Pmean, arterial systolic, diastolic, and mean pressure, respectively; PP, pulse pressure; Rf, total mean resistance; Rc, characteristic impedance; Rs, peripheral resistance; Ca, total compliance; Pes and Pea, ventricular end-systolic and end-diastolic pressure, respectively; CO, cardiac output; EF, ejection fraction, calculated as stroke volume/(end-diastolic volume [Vaved]−volume intercept [Ves]); HR, heart rate; bpm, beats per minute; SW, stroke work; Vaved, end-systolic volume; MV02, myocardial oxygen consumption; PVA, pressure-volume area; EFF, efficiency; α, slope of MV02–PVA relation.

*Upper and lower range estimates for compliance are provided. See “Materials and Methods.”

rapid flow peak, and the square shape of the pressure-volume loop. The next set of examples to the right shows data with flow permitted down both native and Tygon aortas. There was no difference between these data and the waveforms obtained with flow through the native aorta only. This indicates that wave reflections arising from the proximal Dacron conduit or abdominal T-tube connectors were of little influence when flow continued in the native aorta. Clamping the native aorta at the diaphragm while maintaining flow through the Tygon conduit (distal Ao in the figure) resulted in a late-systolic wave reflection, elevating systolic pressures midway into ejection. Repositioning the native clamp proximally (just beyond the Tygon anastomosis) yielded a very different response (far right examples), characterized by a marked widening of the pulse pressure and continuously rising systolic pressures with a late systolic peak. This last configuration achieved the greatest reduction in net vascular compliance and was therefore used for comparison with native aortic flow.

Figure 3 shows typical impedance spectra (modulus and phase) derived from steady-state (left panels) and random-paced (right panels) aortic pressure-flow data for both native (solid line) and Tygon (dashed line) aortas. The moduli for flow through the native aorta rapidly declined to low amplitudes with an early zero-phase crossover. When flow was directed through the Tygon aorta, the modulus decline was more gradual, with a rightward shift of the first modulus minimum and zero-phase crossover frequency. These changes were observed in both the steady-state and random-paced (pseudo-white noise) analyses and are consistent with reduced compliance from the Tygon aorta. Note that the moduli in the random-pacing analysis were normalized to the mean term to better display oscillations at the low frequencies. The coherence for the impedance spectra was near 1.0 up to 12 Hz and then slowly declined, most notably for flow via the Tygon conduit.

These examples were typical of the group changes in vascular loading induced by the switch to a stiff aorta (Table 1). Mean arterial pressure rose slightly from 91.7±20.5 to 102.6±22.5 mm Hg, whereas mean flow (cardiac output) did not significantly change. There was a 20% increase in total mean resistance as well as a rise in characteristic impedance (from 0.27±0.11 to 0.41±0.17 mm Hg·mL⁻¹·sec⁻¹). The most prominent change, however, was a substantial reduction in total vascular compliance. As described in “Materials and Methods,” two compliance estimates were made: an upper value derived from the entire diastolic pressure data and a lower value derived from data beyond the peak of any diastolic pressure oscillations. For the native aorta data, the upper estimate was 1.6 ml/mm Hg, and the lower estimate was
0.7 ml/mm Hg. Despite this range, total compliance with the Tygon aorta was significantly and markedly reduced (0.1–0.2 ml/mm Hg) by 60% to 80%.

The impedance changes with Tygon flow did not result from the small rise in mean arterial pressure. By using loops at slightly reduced preloads, Tygon data were ob-
tained with mean arterial pressure precisely matched to the native aorta flow data (mean pressure difference of +0.3±4.5 mm Hg). However, total peripheral resistance, compliance, and characteristic impedance values obtained from these beats (3.5±1.1 mm Hg·ml⁻¹·sec⁻¹, 0.13±0.12 ml/mm Hg, and 0.44±0.22 mm Hg·ml⁻¹·sec⁻¹, respectively) were nearly identical to those derived from the beats at the slightly higher pressure (Table 1).

Ventricular Systolic Function

Steady-state ventricular function parameters are also provided in Table 1. Increasing the pulsatile load with flow through the Tygon aorta did not significantly change cardiac output, heart rate, ejection fraction, or SW. However, both end-systolic and end-diastolic pressures increased. There was a concomitant small increase in end-systolic volume and Ve (provided in Table 1 as gain-adjusted volume minus Vd). The Ve-Vd change did not reach statistical significance, probably because of the lack of absolute volume calibration as well as statistical uncertainty in Vd estimation.

Pressure–volume relations (and MVO₂–PVA relations) were obtained in nine of the 10 animals. In one animal, preload reduction during flow through the Tygon conduit resulted in diastolic coronary perfusion pressure of ≤40 mm Hg; therefore, the data were not used because of concern for potential cardiac ischemia. Figure 4 displays an example of pressure–volume and MVO₂–PVA data obtained during steady-state reductions in circulating blood volume. ESPVRs (left graphs) were linear and, in this case, showed a slight slope increase with flow through the Tygon aorta. For this example, the SW–Vd relation slope was 59.5 mm Hg for native aorta versus 45.7 mm Hg for Tygon aorta, and dP/dt max at a matched Vd was also slightly less (1,301.7 versus 1,042.5 mm Hg/sec).

Group ESPVR data are provided in Table 2. There was no significant change in slope or offset (Vd) when flow was diverted through the Tygon conduit. This result in part reflected variable responses among experiments. Multivariate regression analysis yielded an average ESPVR of P=6.7 Ve-47.0, where Pe is end-systolic pressure and Ve is end-systolic volume, for native aorta (r²=0.98, n=49) versus Pe=6.98 Ve-42.8 for Tygon aorta (r²=0.93, n=43) (p=NS for slope and Ve comparison between aortas). The Ve is large in these relations primarily from the uncorrected parallel conductance offset of the volume signal. Steady-state dP/dt max matched to the same Ve also did not significantly differ between the two data sets (1,400.6±495 versus 1,702.5±638.7 mm Hg/sec). SW–Vd relations were slightly altered. Although the change was not significant by paired t test of the individual regressions (Table 3), combined multiregression analysis revealed a small but significant change that could be statistically fit by either a pure slope increase (from 74.2±3.0 to 79.8±2.1 mm Hg, p=0.008) or pure Vd (leftward) shift (from

**Figure 4.** Graphs showing end-systolic pressure–volume relations (left panels) and myocardial oxygen consumption (MVO₂)– pressure-volume area (PVA) relations (right panels) for a single dog (No. 5 in Tables 2 and 4) with flow through the native aorta (Ao) versus the Tygon conduit. Volumes were adjusted by subtracting the volume intercept (V0) derived from the native Ao end-systolic pressure–volume relation (ESPVR). There was a slight increase in the ESPVR slope in this example, although overall there was no significant change in this relation. Likewise, the MVO₂–PVA relation was not altered by directing ventricular outflow through the stiff (Tygon) conduit.
62.6±2.6 to 57.8±1.9 ml, p=0.016). Regardless, this effect was small and probably of little physiological importance. Combining the results from all three indexes suggests that there was little to no significant change in systolic contractile function when the heart was made to eject into the stiff aortic conduit.

The above data were obtained with multiple preload volumes at steady state. Since this is an unusual loading pattern for in vivo studies, we also compared pressure-volume relations obtained by more typical transient load interventions (bicaval occlusion). The results (Table 3) were consistent with the steady-state data, displaying no significant change from switching flow between native and Tygon aortas.

**Myocardial Oxygen Consumption and Efficiency**

Flow through the stiff aorta resulted in significant increases in steady-state MVO₂ of more than 30% (Table 1), which was matched by an increase in total pressure-volume work (PVA). An example of MVO₂-PVA relations is also displayed in Figure 4 (right graphs). There was virtually no change in the relation as a consequence of

### Table 2. Systolic Contractility Indexes With Native Versus Tygon Aorta

<table>
<thead>
<tr>
<th>Dog No.</th>
<th>Eₚₑ</th>
<th>V₀⁺</th>
<th>r</th>
<th>Mₛₑ</th>
<th>Vₛₑ⁺</th>
<th>r</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>Native aorta</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>2.8</td>
<td>22.2</td>
<td>0.970</td>
<td>60.8</td>
<td>46.5</td>
<td>0.999</td>
<td>5</td>
</tr>
<tr>
<td>2</td>
<td>6.5</td>
<td>23.6</td>
<td>0.998</td>
<td>73.4</td>
<td>34.9</td>
<td>0.999</td>
<td>5</td>
</tr>
<tr>
<td>3</td>
<td>6.4</td>
<td>61.3</td>
<td>0.997</td>
<td>59.0</td>
<td>70.2</td>
<td>0.999</td>
<td>5</td>
</tr>
<tr>
<td>4</td>
<td>9.0</td>
<td>37.5</td>
<td>0.994</td>
<td>70.2</td>
<td>47.0</td>
<td>0.990</td>
<td>6</td>
</tr>
<tr>
<td>5</td>
<td>2.8</td>
<td>89.6</td>
<td>0.994</td>
<td>61.7</td>
<td>114.2</td>
<td>0.997</td>
<td>7</td>
</tr>
<tr>
<td>6</td>
<td>7.9</td>
<td>84.5</td>
<td>0.875</td>
<td>105.3</td>
<td>98.0</td>
<td>0.992</td>
<td>5</td>
</tr>
<tr>
<td>7</td>
<td>6.0</td>
<td>25.9</td>
<td>0.981</td>
<td>68.9</td>
<td>38.2</td>
<td>0.996</td>
<td>5</td>
</tr>
<tr>
<td>8</td>
<td>11.7</td>
<td>33.2</td>
<td>0.912</td>
<td>99.2</td>
<td>42.2</td>
<td>0.999</td>
<td>4</td>
</tr>
<tr>
<td>9</td>
<td>7.6</td>
<td>58.6</td>
<td>0.991</td>
<td>65.4</td>
<td>69.1</td>
<td>0.992</td>
<td>7</td>
</tr>
<tr>
<td>Mean</td>
<td>6.74</td>
<td>48.48</td>
<td>0.968</td>
<td>73.77</td>
<td>62.26</td>
<td>0.996</td>
<td>5.4</td>
</tr>
<tr>
<td>SD</td>
<td>2.82</td>
<td>26.05</td>
<td>0.044</td>
<td>16.88</td>
<td>28.05</td>
<td>0.004</td>
<td>1.0</td>
</tr>
<tr>
<td>Tygon aorta</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>3.1</td>
<td>29.8</td>
<td>0.944</td>
<td>46.6</td>
<td>46.8</td>
<td>0.919</td>
<td>7</td>
</tr>
<tr>
<td>2</td>
<td>10.0</td>
<td>20.7</td>
<td>0.976</td>
<td>90.2</td>
<td>32.1</td>
<td>0.983</td>
<td>4</td>
</tr>
<tr>
<td>3</td>
<td>5.9</td>
<td>46.8</td>
<td>0.653</td>
<td>93.2</td>
<td>67.1</td>
<td>0.932</td>
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</tr>
<tr>
<td>4</td>
<td>15.3</td>
<td>48.7</td>
<td>0.935</td>
<td>142.3</td>
<td>49.8</td>
<td>0.931</td>
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</tr>
<tr>
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<td>102.7</td>
<td>0.970</td>
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<td>122.3</td>
<td>0.995</td>
<td>7</td>
</tr>
<tr>
<td>6</td>
<td>5.6</td>
<td>85.3</td>
<td>0.803</td>
<td>78.1</td>
<td>103.5</td>
<td>0.920</td>
<td>5</td>
</tr>
<tr>
<td>7</td>
<td>7.8</td>
<td>20.8</td>
<td>0.840</td>
<td>86.3</td>
<td>36.5</td>
<td>0.760</td>
<td>4</td>
</tr>
<tr>
<td>8</td>
<td>4.6</td>
<td>10.8</td>
<td>0.930</td>
<td>84.2</td>
<td>36.5</td>
<td>0.999</td>
<td>4</td>
</tr>
<tr>
<td>9</td>
<td>8.4</td>
<td>56.4</td>
<td>0.966</td>
<td>67.8</td>
<td>70.4</td>
<td>0.999</td>
<td>5</td>
</tr>
<tr>
<td>Mean</td>
<td>7.14</td>
<td>46.88</td>
<td>0.891</td>
<td>81.28</td>
<td>62.78</td>
<td>0.938</td>
<td>4.78</td>
</tr>
<tr>
<td>SD</td>
<td>3.81</td>
<td>30.90</td>
<td>0.107</td>
<td>29.27</td>
<td>31.66</td>
<td>0.075</td>
<td>1.39</td>
</tr>
</tbody>
</table>

ESPVR, end-systolic pressure-volume relation; SW, stroke work; Vₑₑ, end-diastolic volume; Eₚₑ, slope value for ESPVR; V₀⁺, volume intercept for ESPVR; r, correlation coefficient; Mₛₑ, slope value for the SW–Vₑₑ relation; Vₛₑ⁺, volume intercept for SW–Vₑₑ relation; n, number of points per regression.

**Table 3. Comparison of Indexes of Cardiac Contractility Derived From Transient Preload Reduction (Simultaneous Inferior Vena Cava and Superior Vena Cava Occlusion) Versus Steady-State Changes in Mean Circulating Volume**

<table>
<thead>
<tr>
<th></th>
<th>Eₚₑ (mm Hg/ml)</th>
<th>Mₛₑ (mm Hg)</th>
<th>dP/dtmax, at matched Vₑₑ (mm Hg/sec)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Native aorta</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acute</td>
<td>5.47±2.3</td>
<td>65.35±11.11</td>
<td>1,471.41±483.14</td>
</tr>
<tr>
<td>Steady state</td>
<td>6.44±3.0</td>
<td>77.08±19.1</td>
<td>1,400.56±495.48</td>
</tr>
<tr>
<td>Tygon aorta</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acute</td>
<td>7.80±5.1</td>
<td>76.0±24.39</td>
<td>1,374.71±493.25</td>
</tr>
<tr>
<td>Steady state</td>
<td>6.21±2.2</td>
<td>88.08±26.1</td>
<td>1,702.54±638.69</td>
</tr>
</tbody>
</table>

Eₚₑ, slope of end-systolic pressure–volume relation; Mₛₑ, slope of stroke work–end-diastolic volume relation; Vₑₑ, end-diastolic volume.

All the indexes demonstrated statistically similar results from both acute-load vs. steady-state load alteration methodologies. The comparisons were not statistically significant by paired t test with Bonferroni correction for multiple testing.
switching from the native to the Tygon aorta. This was found in nearly all the animals (Table 4). MVo2-PVA relations were also analyzed by multiple regression, which revealed an insignificant effect of aorta type on overall regression variance \((p=0.71)\). The mean pooled regression relation based on all 92 points from the 18 runs was as follows: MVo2=2.04\times10^{-5} \cdot \text{PVA}+0.019 \quad (p<0.0001, \text{multiple } r=0.993, \text{SEE}=0.0025).

Steady-state myocardial efficiency was determined in two ways. One was the ratio of ventricular external work to MVo2 (both converted to joules). Increased vascular stiffness reduced this efficiency from 22.8% to 17.6% \((p=0.052)\). This ratio, however, is load dependent, whereas the inverse slope of the MVo2-PVA relation provides a relatively load-insensitive measure of chemomechanical efficiency.\(^{10}\) Switching to the Tygon conduit did not alter this latter efficiency \((35.5\pm11.2\% \text{ for native aorta versus } 34.4\pm9.13\% \text{ for Tygon aorta, } p=NS)\).

Although cardiac efficiency was little altered, the energetic costs to the heart for delivering a given SV (or cardiac output) were significantly increased when flow was directed via the stiff conduit. An individual example of the relation between SV and MVo2 is shown in Figure 5A. SV and MVo2 were linearly correlated (mean correlation coefficient, 0.93\pm0.06) in both native and Tygon aortic flow conditions, but MVo2 was greater at any given SV (upward shift of this relation) when flow was directed through the stiff aorta.

Group results are shown in Figure 5B. To graphically combine data, linear fits to the native aorta data \((\text{MVo2}=a \cdot \text{SV}+b, \text{where } a \text{ is the slope and } b \text{ is the y intercept})\) were used to generate a new variable, \(\text{SV}'=a' \cdot \text{SV}+b'\), and the same equation was then applied to both native and Tygon aorta data for each animal respectively. The MVo2-SV' points fell about the iden-
tity line for native aorta data, whereas the Tygon data were normalized to this control relation and thus demonstrated relative shifts due to the stiffer vasculature (Figure 5B). The group results were similar to the result shown in the individual example of Figure 5A. Multiregression analysis of the raw data yielded the mean relation

\[ \text{MVO}_2 = -0.034 + 6.13 \times 10^{-3} \cdot SV + 1.11 \times 10^{-2} \cdot G \]

where G is a grouping variable (G = 0 for native aorta, G = 1 for Tygon aorta), with all coefficients significant at \( p < 0.001 \). Thus, for a typical SV (15 ml, from mean cardiac output/heart rate; see Table 1), flow through a stiff vasculature required 20% more MVO₂. The range of increased MVO₂ spanned 15–41%, depending on the baseline SV.

**Discussion**

This study investigated the effects of left ventricular ejection into a stiff vascular system on in vivo systolic mechanics and energetics. Aortic input impedance and central arterial pressure and flow analyses demonstrated that redirecting flow from the native thoracic aorta through a stiff conduit markedly lowered total compliance, increased characteristic impedance, and only slightly changed total mean resistance. These loading changes are similar to those observed in human aging studies. Interestingly, this marked vascular load change did not significantly alter indexes of chamber systolic function or efficiency based on analysis of pressure–volume and MVO₂–PVA relations. However, the cardiac energetic cost of delivering a given SV increased by 20–40%. This suggests a mechanism whereby vascular stiffening in humans may yield little functional decrement at rest but limit reserve capacity under stress conditions.

**Myocardial Energetics**

Two prior in vivo studies reported MVO₂ changes after primary reductions in arterial compliance, one directly by means of a left and right heart bypass preparation and the other by radiolabeled microspheres. In both instances, the results were variable and inconclusive. To our knowledge, the present study provides the first demonstration that ejection into a stiff vasculature results in a significant change in MVO₂. There are several potential sources for the observed increase including 1) a positive inotropic effect from the increased arterial load, analogous to an Anrep effect, 2) an intrinsic reduction in the efficiency of chemomechanical energy transduction, or 3) a corresponding increase in total cardiac work. The present results favor the latter mechanism since none of the indexes of systolic function or chamber efficiency were significantly altered by the switch to the stiff aortic conduit. Instead, the increase in MVO₂ was appropriate to the elevated total PVA.

As suggested by isolated heart data, it is unlikely compliance reduction itself explained the increase in resting PVA (and MVO₂). Rather, in vivo compensatory mechanisms such as a small rise in \( V_{ed} \) likely played an important role. \( V_{ed} \) change could not be precisely determined from the present data, since the volume offset (parallel conductance) of the catheter signal was not measured. However, end-diastolic pressure rose significantly (Table 1), and there was a suggestion that \( V_{ed} - V_e \) also increased. Another factor that could have contributed to the PVA increase was the small elevation in mean total resistance. The exact cause for this change is unclear but is likely related in part to increased reactive load from the Tygon tubing and its connections, indicated by the rise in characteristic impedance.

Two prior investigations of the relation between MVO₂ and PVA, most conducted in isolated canine hearts. First described by Suga and colleagues, this relation has been repeatedly found to be little influenced by altered ejection history or loading (even when hearts are filled during systole). However, we recently reported that increasing afterload resistance in isolated hearts (at constant compliance and characteristic impedance) results in a lowering of efficiency (greater MVO₂–PVA relation slope) as well as reduction in ESPVR slope. Central to these phenomena were changes in the extent of ejection, or the effective ejection fraction (SV/[\( V_{ed} - V_e \)]), that accompanied altered resistance load. There was a direct correlation between decreased effective ejection fraction and lowered efficiency. In the present study, the largely pulsatile loading changes induced by the Tygon conduit resulted in little change in the effective ejection fraction (from 50% to 43%, \( p = \text{NS} \)). This is consistent with the observed insignificant effects on the MVO₂–PVA relation.

The relation between SV and MVO₂ is not an “efficiency” per se, in that SV cannot be expressed in energy units. However, it is a relevant measure of the cardiac costs of delivering a given systemic flow when the vasculature is stiffened, and the present data showed that this could be substantial. Despite additional MVO₂ requirements, systolic function was not depressed as might result from an O₂ supply/demand imbalance. However, the present study was conducted in normal canine hearts. Human diseases in which arterial stiffening occurs are often associated with coronary artery stenoses (from atherosclerosis) or cardiac hypertrophy (from hypertension or arteriosclerosis). It remains possible that under these conditions, in which the baseline cardiac supply/demand balance is already compromised, vascular stiffening could have far more profound effects.

Two other aspects of the MVO₂–SV relation deserve comment. It was obtained by varying preload at a presumed fixed afterload impedance. Certainly, a very different relation would result if afterload resistance were primarily altered. The similarity of pressure-volume loop shape despite preload reduction (Figure 4) indicates that the dominant characteristics of impedance loading for both native and Tygon aortas were fairly constant during volume reduction. However, some load dependence of both compliance and reflected waves was certainly present. This likely contributed to the apparent negative offset constant for the MVO₂–SV relation (i.e., zero MVO₂ at a positive SV). This result based on linear extrapolation suggests that the true relation is nonlinear at low volumes. In addition to volume (or mean pressure) dependence of vascular impedance load, ESPVR curvilinearity could play a role.

**Systolic Function**

The ESPVR was first described to be little influenced by varying impedance loads; however, subsequent
studies have revealed influences of ejection history. Several investigators described negative effects of ejection\textsuperscript{31–33} related to the overall extent, rate, and pattern of shortening. More recent isolated heart studies\textsuperscript{19,34,35} have reported positive effects of ejection. In vivo studies have reported either leftward ESPVR shifts with little slope change accompanying afterload resistance increase\textsuperscript{55} or significant differences in slope depending on load intervention\textsuperscript{56} (preload versus afterload). No prior study has compared in vivo ESPVRs for which, primarily, the pulsatile components of aortic input impedance rather than mean peripheral resistance were increased.

The lack of significant change in the ESPVR in the present study is remarkable and suggests that the pattern of ejection (i.e., the rate of flow and pressure changes) may be a less critical determinant of in vivo systolic function than is the total extent or average rate of ejection. This result is further supported by the lack of change in the SW–\(V_e\) relation or \(dP/dt_{max}\) at matched \(V_e\). While the SW relation is anticipated to change with marked alterations in afterload impedance,\textsuperscript{37} this, if anything, would decrease the slope. Yet the average direction of this slope was toward a small increase. It is also worth noting that SV (and thus cardiac output) was little influenced despite the marked reduction in compliance and increase in characteristic impedance. This is consistent with data reported in isolated canine hearts ejecting into a simulated three-element Windkessel model\textsuperscript{38} and points out the relatively minor impact of high-frequency components of the impedance spectrum on integrated parameters such as SV.

Comparison With Prior Models of Arterial Stiffening

Experimentally altering the elastic properties of the thoracic aorta is difficult, and results may partially depend on the method used. A classic study by Salisbury et al\textsuperscript{22} used an entirely artificial hydraulic Windkessel model in an isolated in situ heart preparation. Unfortunately, these preparations often introduce bizarre wave reflections, so that the “arterial” waveforms are not realistic, even though calculations of mean resistance or capacitance may be reasonable. Urschel et al\textsuperscript{20} bypassed only the descending aorta (by a glass tube), leaving a substantial portion of the total vascular capacitance (the proximal ascending aorta and aortic root) intact. This same limitation applies to the study of O’Rourke,\textsuperscript{21} who “stiffened” the descending aorta by attaching Lucite to the outer surface. In contrast, Randall et al\textsuperscript{23} placed an endotracheal tube within the lumen of the ascending aorta, and by inflating or deflating the cuff balloon, directed flow either around or solely within the tube. Whereas this approach reduced the extent of residual aortic compliance (reduction varied from 30% to 60%), it also introduced a sudden diameter change when the cuff was inflated and thus an acute impedance mismatch.

The present preparation was certainly invasive, yet by attention to a number of key factors, reasonably physiological data were obtained. The Dacron anastomosis was sutured to the ascending aorta so that much of the thoracic aortic compliance was bypassed. The experiment was designed so that the real native aorta and the stiff Tygon aorta could be studied in the same animal. Finally, circumferential pressures, cardiac outputs, and systolic and diastolic function were maintained in a physiologic range. Evidence supporting the realistic nature of the preparation is found in comparison to human data obtained in young versus old hypertensive subjects (Figure 6). These clinical examples were determined by similar conductance catheter methods, with preload transiently altered by temporarily occluding the inferior vena cava with a balloon catheter. The young patient displays a “square” shape pressure–volume loop, whereas the older hypertensive subject displays a marked late systolic peaking of the pressure–volume loop. Below this figure is an example from the present canine study with native versus Tygon aortas. The similarities are striking, suggesting that, despite its invasiveness, the present model provides a clinically relevant method for assessing the influence of vascular stiffness on ventricular mechanics and energetics.

Limitations

It has been suggested that, in addition to compliance changes, alteration in the timing and intensity of wave reflections may contribute to impaired cardiac function\textsuperscript{6} with ejection into a stiff vasculature. Although the aortic bypass preparation in the present study primarily altered aortic compliance, increased wave reflections could also have played a role. However, it is difficult, if not impossible, to separately control or quantify these two factors in vivo. Diastolic reflections, which were often present in the native aorta data, complicated compliance estimations. Our results using the area method of Liu et al\textsuperscript{27} likely overestimated total vascular compliance when the entire diastolic period

\begin{figure}
\centering
\includegraphics[width=0.8\textwidth]{figure6}
\caption{Left ventricular (LV) pressure–volume loops obtained from a normal young patient (aged 19 years) and an older hypertensive patient (aged 53 years) compared with the canine bypass model of the present study with flow through the normal versus the Tygon aorta. Flow through the native canine aorta produced data very similar to that typically seen in young human subjects with compliant vasculatures. In contrast, the older patient demonstrates late systolic peaking, which was well simulated by the flow through the Tygon conduit.}
\end{figure}
was used and underestimated it when only late declining pressures were analyzed. However, even the lower estimate was somewhat greater than previously report-
ed.23,30 Two other factors that likely contributed to this are the large dog size used to facilitate the surgical preparation and the effects of sympathetic withdrawal due to hexamethonium-induced autonomic blockade.

An important caveat to this study is the fact that the switch to a stiff conduit was done acutely. In states of chronic vascular stiffening, left ventricular hypertrophy may have an important influence on cardiac efficiency. The increase in M\textsubscript{VO\textsubscript{2}} may be affected by concomitant left ventricular hypertrophy–related changes in contractility. Furthermore, the extent to which increased oxygen demand imposed by a stiff vasculature can be met by augmented coronary perfusion is anticipated to be more severely limited with left ventricular hypertrophy.

In isolated heart studies, the right ventricle is vented, and as unloaded independent of the left ventricle, making the calculation of left ventricular M\textsubscript{VO\textsubscript{2}} more direct. Since we did not use a right heart bypass in situ, the amount of left heart oxygen use was estimated by the ratio of left ventricular to total ventricular mass. This assumes that the net production of \textsubscript{O\textsubscript{2}} consumption between ventricles remains constant independent of volume loading. Errors in this assumption would result in slightly different slopes and offsets for the M\textsubscript{VO\textsubscript{2}}–PVA relations from those obtained in isolated hearts, although the present values are consistent with several prior studies.16–18 With respect to the change from native to Tygon aorta, the Tygon aorta might have loaded the left heart more than the right, making the energetic costs somewhat greater than we calculated. This effect could slightly lower the efficiency during ejection into the Tygon aorta and shift the M\textsubscript{VO\textsubscript{2}}–SV relation upward.

Finally, we did not estimate the parallel conductance offset of the volume catheter signal, because this adds complexity and potential instability to the preparation, and it was not required for most aspects of the mechanical or energetic analyses. It remains possible, however, that small volume shifts in the ESPVR or SW–V\textsubscript{e}d relations were obscured by changes in the parallel offset. Countering this are the similar results obtained from the transient preload reduction data (Table 3), which were obtained quickly and temporally closer together (native versus Tygon aorta), making parallel conductance change less likely.

Summary

Vascular changes with aging and hypertension have been extensively studied in humans, and there is continued speculation that vascular stiffening poses adverse conditions for the heart. The present study shows that in normal hearts, the effects on contractile function and efficiency are minimal. These data mark an important step toward understanding how the aging vasculature may change cardiac performance and energetics. As demonstrated in Figure 6, the model reproduces many of the pressure–volume characteristics observed in humans, and the compliance reduction achieved is consistent with estimates from human aging studies. Future investigations will need to examine specifically the interactions between pulsatile loading and diseased hearts, where clinical limitations are most likely to be observed.

Acknowledgments

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